



Technical Manual

Human Mumps Virus IgM ELISA Kit

- Catalogue Code: HDES0016
- Antibody ELISA Kit
- Research Use Only

1. Test principle

This ELISA kit uses Capture-ELISA as the method to detect the MuV-IgM in human serum. The ELISA Microtiter plate provided in this kit has been pre-coated with Mouse-anti-human IgM (μ chain). Samples are added to the ELISA Microtiter plate wells and the IgM antibody in which will be captured. Free components are washed away (including the specific IgG antibody). The “MuV antigen + anti- MuV -HRP conjugated antibody” is added to each well, the MuV -IgM antibody in captured IgM will specific bind to the MuV antigen + anti- MuV -HRP conjugated antibody. The TMB substrate is added after washing to initiate the color developing reaction. The presence of HV-IgM can be determined according to the OD value after colorimetric assay with the Micro-plate Reader.

2. Kit components

Item	Specification
ELISA Microtiter plate	96 wells
Positive Control	1 mL
Negative Control	1 mL
HRP Conjugate	12 mL
Sample Diluent	12 mL
20×Concentrated Wash Buffer	50 mL
Substrate Reagent A	6 mL
Substrate Reagent B	6 mL
Stop Solution	6 mL
Plate Sealer	3 pieces
Sealed Bag	1
Manual	1 copy

3. Experimental instrument

- Micro-plate Reader with 450 nm wavelength filter or dual-wavelength (450/630 nm)
- High-precision transferpettor, EP tubes and disposable pipette tips
- 37° C Incubator or water bath
- Deionized water
- Absorbent paper
- Loading slot for Wash Buffer

4. Requirements of sample

1. Human serum can be used as detected sample. Fresh collected serum samples should be fully centrifuged, then take clear liquid for test. The suspended fibrous protein may cause a false positive result if not fully precipitated. Avoid of samples with hyperlipidemia (triglyceride ≥ 20 g/L), hemolysis (hemoglobin ≥ 10 g/L) or jaundice (bilirubin ≥ 0.2 g/L). Obviously contaminated samples can't be detected.
2. Do not use heated inactivated samples. Heat inactivation will degrade antibodies.
3. Samples can be stored at 2~8° C for one week. If samples not tested in a week, store them at -20° C and avoid freeze-thaw cycles.
4. Bring all reagents to room temperature (18~25° C) for more than 30 min before use. Freezing samples should be mixed fully before test.

5. Assay procedure

Bring all reagents to room temperature for 30 min. Dilute the 20×Concentrated Wash Buffer for 20 times with distilled water.

1. Add sample:

- (1) Take out Micro-plate and mark it, reserve 1 well for blank control (empty), 3 wells for negative control, 2 wells for positive control (100 μ L of control serum for each well). (Blank well is not necessary for dual-wavelength detection)
- (2) Dilute the tested Serum with Sample Diluent at 1:10 into sample well (add 100 μ L of Sample Diluent and add 10 μ L of serum sample), mix fully.
- (3) Gently tap the plate to ensure thorough mixing.

2. Incubate: Cover the ELISA plate with sealer. Incubate for 30 min at 37° C.

3. Wash: After incubation, remove the plate sealer and aspirate the liquid of each well. Repeat the washing procedure for 5 times with wash buffer and immerse for 30-60 sec each time.

4. HRP conjugate: Add 100 μ L of HRP Conjugate Working Solution to each well except the blank control well.

5. Incubate: Cover the ELISA plate with sealer. Incubate for 30 min at 37° C.

6. Wash: After incubation, remove the plate sealer and aspirate the liquid of each well. Repeat the washing procedure for 5 times with wash buffer and immerse for 30-60 sec each time.

7. Add substrate: Add 50 μ L of Substrate Reagent A and 50 μ L of Substrate Reagent B to each well. Gently tap the plate to ensure thorough mixing. Cover with a new plate sealer. Incubate for 15 min at 37° C in dark.

8. Stop reaction: Add 50 μ L of Stop Solution to each well, gently tap the plate to ensure thorough mixing.

9. OD Measurement: Set the micro-plate reader wavelength at 450 nm (it is recommended

to set the dual wavelength at 450 nm/630 nm) to detect A value of each well. Blank well is not needed when using dual wavelength 450 nm/630 nm for detection.

6. Reference value

1. Result analysis

- (1) Use each test result independently. Determine the result according to the Cut Off value.
- (2) Calculate the Cut Off: $\text{Cut Off(C.O)} = 0.10 + \text{negative control (NC) average A value}$ (when NC average $A_{450} < 0.05$, calculate at 0.05; while NC average $A_{450} \geq 0.05$, calculate at the actual value).

2. Quality control

- (1) Blank well (just chromogenic agent and stop solution) absorbance ≤ 0.08 .
- (2) Positive control (PC) $A_{450} > 0.80$.
- (3) Negative control (NC) $A_{450} < 0.10$.

The experimental result is valid if quality control is valid.

3. Determination of results

- (1) Positive result: Sample absorbance $\geq$ Cut Off.
- (2) Negative result: Sample absorbance $<$ Cut Off.

7. Interpretation of results

1. Negative result indicates there is no MuV-IgM antibody detected in samples, while positive result means the opposite.
2. The positive result of MuV -IgM antibody is an important index of MuV acute infection.

8. Limitations of test method

1. All high sensitivity immune experiment system exists potential non-specificity. Therefore, unacceptable positive results may be caused by biological false positive of ELISA method.
2. In the early stage of infection, IgM did not occur or has a low titer, and these situations will lead to negative results. It is recommended to remind the patients to recheck within 7-14 days. Make a parallel detection of the last sample to confirm whether there is seroconversion or titer elevation.
3. The reference value of serum antibody detection of patients with impaired immune function or patients receiving immunosuppressive therapy is limited.
4. Any positive result should be determined combined with clinical information.

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