



Technical Manual

ERY (Erythromycin) ELISA Kit

- **Catalogue Code: FSES0055**
- **Competitive ELISA Kit**
- **Research Use Only**

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1. Key features and Sample Types

Sensitivity:

0.1 ppb (ng/mL)

Assay Procedure:

25°C, 30 min~30 min~15 min

Detection Limit:

Raw milk, Egg - 2 ppb; Urine, Serum, Muscle - 5 ppb;
Muscle (fish, shrimp) - 10 ppb.

Cross Reactivity:

Erythromycin - 100%

Spiramycin, Valnemulin, Lincomycin, Kanamycin, Tilmicosin, Doramectin, Ivermectin,
Eprinomectin, Moxidectin, Abamectin, Tylosin - < 0.1%

Sample Recovery rate:

90%±30%.

Storage:

2-8°C for 6 months.

Expiry:

See Kit Label

2. Storage

Store the kit at 2~8°C. Do not freeze any test kit components.

Return any unused microwells to their original foil bag and reseal them together with the desiccant provided and further store at 2-8°C.

3. Test Principle

This kit uses a Competitive-ELISA method. It can detect Erythromycin (ERY) in samples, such as muscle, serum, etc. This kit is composed of ELISA Microtiter plate, HRP conjugate, antibody working solution, standard and other supplementary reagents. The microtiter plate in this kit has been pre-coated with coupled antigen. During the reaction, ERY in the samples or standard competes with coupled antigen on the solid phase supporter for sites of anti-ERY antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each microtiter plate well, and Substrate Reagent is added for color development. There is a negative correlation between the OD value of samples and the concentration of ERY. The concentration of ERY in the samples can be calculated by comparing the OD of the samples to the standard curve.

4. Kit Contents

Each kit contains reagents for 96 assays including:

No.	Component	96-WellKit
1	ELISA Microtiter Plate	96 wells
2	Standards	1 mL each (0 ppb, 0.1 ppb, 0.3 ppb, 0.9 ppb, 2.7 ppb, 8.1 ppb)
3	HRP Conjugate	12 mL
4	Antibody Working Solution	10 mL
5	Substrate Reagent A	6 mL
6	Substrate Reagent B	6 mL
7	Stop Solution	6 mL
8	20xConcentrated Wash Buffer	25 mL
9	Tissue Diluent	50 mL
10	Plate Sealer	3 pieces
11	Sealed Bag	1 piece
12	Manual	1 copy

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution.

Additional materials required:

Other materials required but not supplied

- **Instrument:** Microplate reader, Vortex mixer, Centrifuge, Balance (sensitivity 0.01 g).
- **Micropipette:** Single channel (20-200 μ L, 100-1000 μ L), Multichannel (30-300 μ L).
- **Reagents:** Methanol, N, N-Dimethylformamide (DMF), NaCl, NaOH, Concentrated HCl.

5. Experimental Preparation

Bring all reagents and samples to room temperature before use.

Open the microplate reader in advance, preheat the instrument, and set the testing parameters

1. Sample pre-treatment Notice:

Experimental apparatus should be clean, and the pipette should be disposable to avoid cross- contamination during the experiment.

2. Solution preparation

Please prepare solution according to the number of samples. Don't use up all components in the kit at once!

Solution 1: 6% DMF Solution (*for swine, serum bovine sample*)

Dilute 6 mL of **N, N-Dimethylformamide** to 100 mL with deionized water, mix fully.

Solution 2: 2% NaCl Solution (*for eggs sample*)

Dissolve 2 g of **NaCl** to 100 mL with deionized water, mix fully.

Solution 3: 0.05 M NaOH Solution (*for livestock sample*)

Dissolve 1 g of **NaOH** to 500 mL with deionized water, mix fully.

Solution 4: 0.1 M HCl Solution (*for raw milk, livestock sample*)

Dilute 0.86 mL of **HCl** to 100 mL with deionized water, mix fully.

Solution 5: Wash Buffer

Dilute **20×Concentrated Wash Buffer** with deionized water.

(**20×Concentrated Wash Buffer** (V): Deionized water (V) = 1:19).

Solution 6: Sample Diluent (*for raw milk, fish, shrimp sample*)

Dilute **20×Concentrated Wash Buffer** with deionized water.

(**20×Concentrated Wash Buffer** (V): Deionized water (V) = 1:99).

3. Sample pre-treatment procedure

Targets may be distributed unevenly, resulting in no detection. To avoid this, ensure to take sufficient samples when sampling.

3.1 Pre-treatment of raw milk sample:

1. Take 1 mL of the fresh raw milk into a centrifuge tube, and add 100 μ L of **0.1 M HCl Solution** (Solution 4). Vortex for 1 min to mix thoroughly. Centrifuge for 10 min at 4000 r/min.
2. Remove the upper layer of fat, take 100 μ L of the solution in intermediate layer into another centrifuge tube, and add 900 μ L of **Sample Diluent** (Solution 6). Vortex for 10 s to mix thoroughly.
3. Take 20 μ L for detection.

Note: Sample dilution factor: 10, detection limit: 2 ppb

3.2 Pre-treatment of eggs sample:

1. Take 50 μL of homogenate eggs sample into a centrifuge tube, add 950 μL of **2% NaCl Solution** (Solution 2). Vortex for 10 s to mix thoroughly.
2. Take 20 μL for detection.

Note: Sample dilution factor: 20, detection limit: 2 ppb

3.3 Pre-treatment of urine (swine)*, serum (bovine) sample:

(*Data validated in swine urine but pre-treatment can be applied for urine samples of multiple species.)

1. Sample should be mixed thoroughly before test.
2. Take urine sample into centrifuge for 5 min at 4000 r/min.
3. **Urine sample:** Take 100 μL of the supernatant of urine into another centrifuge tube after the centrifuge.
Serum sample: Take 100 μL of homogenate serum into another centrifuge tube after the centrifuge.
4. Add 900 μL of **6% DMF Solution** (Solution 1) and vortex for 5min to mix thoroughly.
5. Take 20 μL for detection.

Note: Sample dilution factor: 10, detection limit: 5 ppb

3.4 Pre-treatment of muscle (livestock) sample:

1. Weigh 1 ± 0.05 g of homogenate sample into a 50 mL of centrifuge tube. Add 4 mL of **0.05 M NaOH Solution** (Solution 3), and vortex for 3 min.
2. Centrifuge for 5 min at 4000 r/min.
3. Take 100 μL of supernatant to another centrifuge tube, add 375 μL of **Tissue Diluent**, then add 25 μL of **0.1 M HCl Solution** (Solution 4), and vortex for 10 s thoroughly.
4. Take 20 μL for detection.

Note: Sample dilution factor: 20, detection limit: 5 ppb

3.5 Pre-treatment of muscle (fish, shrimp) sample:

1. Weigh 2 ± 0.05 g of homogenate sample into a 50 mL of centrifuge tube.
2. Add 1.2 mL of deionized water, then add 2.8 mL of **Methanol**, vortex for 1 min.
3. Centrifuge for 10 min at 4000 r/min.
4. Take 200 μL of the supernatant to another centrifuge tube, and add 800 μL of **Sample Diluent** (Solution 6), fully vortex for 30 s.
5. Take 20 μL of supernatant for detection.

Note: Sample dilution factor: 15, detection limit: 10 ppb

6. Assay Procedure

Bring all reagents and samples to room temperature (25°C) before use. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. The unused ELISA Microtiter plate should be sealed as soon as possible and stored at 2~8°C.

1. **Number:** number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells. **Standard and Samples must be tested in duplicate.**
2. **Add Sample:** add 20 µL of **Standard or Sample** per well, add 80 µL of **Antibody Working Solution** into each well. Gently vortex for 10 s to mix thoroughly and cover the plate with sealer. Incubate min at 25°C for 30 min away from direct sunlight.
3. **Wash:** uncover the sealer carefully, remove the liquid in each well. Immediately add 260 µL of **Wash Buffer** (Solution 5) to each well and wash. Repeat the wash procedure for 4 times, 30 s intervals/time. Invert the plate and pat it against absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **HRP Conjugate:** add 100 µL of **HRP Conjugate** into each well, and incubate at 25°C for 30 min.
5. **Wash:** Repeat step 3 for washing.
6. **Colour Development:** add 50 µL of **Substrate Reagent A** to each well, and then add 50 µL of **Substrate Reagent B**. Gently vortex for 15 s to mix thoroughly. Incubate for 15 min at 25°C away from direct sunlight (The reaction time can be extended according to the actual colour change).
7. **Stop Reaction:** add 50 µL of **Stop Solution** to each well, vortex gently for 10 s to mix thoroughly.
8. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. This step should be finished in 5 min after stop reaction.

7. Data Analysis

1. **Absorbance (%) = $A/A_0 \times 100\%$**

A: Average absorbance of standard or sample

A₀: Average absorbance of 0 ppb Standard

2. **Drawing and calculation of standard curve**

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. **If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.**

For this kit, it is more convenient to use professional analysis form for accurate and fast analysis of batch samples.

8. Notes

1. The overall OD value will be lower when reagents have not been brought to room temperature before use or room temperature is below 25°C.
2. If the wells turn dry during the washing procedure, it will lead to bad linear standard curve and poor repeatability. Operate the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. ELISA Microtiter plate should be covered by plate sealer. Avoid the kit to strong light.
5. **Each reagent is optimized for use in the FSES0055. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other FSES0055 with different lot numbers.**
6. Substrate Reagent should be abandoned if it turns blue colour. When OD value of standard (concentration: 0) < 0.8 unit (A450nm < 0.8), it indicates the reagents are deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.
8. As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.
9. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
10. If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary.
11. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.

Notes:

Assay Genie 100% money-back guarantee!

If you are not satisfied with the quality of our products and our technical team cannot resolve your problem, we will give you 100% of your money back.

Contact Details



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